

SUPPLEMENTARY MATERIAL

Brief Discussion of Fossils not Used for Calibration

Several fossils have not been included in the present study, because of their doubtful taxonomic affinities and poor stratigraphic constraints. Relevant examples include:

†*Viviparus* (?) *albascopularis* was described from the Australian Wallumbilla Formation (Early Cretaceous, Aptian, c. 125-112 Myr) by Etheridge (1902) representing the earliest record of a freshwater gastropod from Australia. Kear et al. (2003) later re-assessed this fossil and placed it within *Notopala* based on comparative morphology and morphometry. However, the single available specimen lacks its protoconch and lacks an intact aperture and thus several diagnostic traits cannot be assessed. Moreover, it has been found in marine deposits and was therefore reworked.

Fossils described by Hislop (1860) from the Indian Deccan Plateau Formation (Late Cretaceous, Maastrichtian, 70-65 Myr) have tentatively been assigned to the genus *Bellamyia* (Hartman et al. 2008). Whereas these shells resemble the general habitus of smooth-shelled viviparids, they are much smaller than is typical for the family. Moreover, their preservation does not allow to evaluate diagnostic features which renders their placement within the Viviparidae is questionable.

Fossil viviparids have been also found in South America, namely †*Viviparus wichmanni* from the Late Cretaceous of Argentina (Doello-Jurado 1922) and †*Paludina araucana* from the Tertiary of Chile (Philippi 1887). However, their assignment to the Viviparidae remains questionable due to non-characteristic descriptions as already indicated by Prashad (1928). Moreover, the colonization of South America out of Laurasia during the Cretaceous is very unlikely according to Prashad (1928) and our palaeogeographical reconstructions (Fig. S6).

The North American *Tulotomops* (only known from the Upper Cretaceous) and *Lioplacodes* cannot be unambiguously assigned to Viviparidae.

Remarks on Habitat Types for the Five Fossils Used to Time-Calibrate the Phylogeny

Inference of the habitat of fossil taxa requires accurate knowledge of the depositional environment and relevant taphonomic processes. Moreover, the local occurrence of fossil assemblages may only poorly reflect the total geographical range of the taxon in question. We therefore took a conservative approach when inferring habitat types for fossil taxa.

†*Viviparus langtonensis* (CP1) – **habitat type: uncertain ('LeA/LoA')**

Freshwater-brackish (lagoon-like) habitat

†*Campeloma harlowtonense* (CP2) – **habitat type: probably lotic ('LoA')**

According to (Yen 1950), the associated fauna suggests that the origin of the Kootenai Formation was likely fluvatile. He also compared the Kootenai mollusc assemblage with the recent mollusc fauna of the Great Falls of the Potomac River, near Washington, D.C., USA and found several similarities. Although Yen (1950) further notes that a lacustrine origin should also be considered, a riverine origin seems most likely.

†*Margarya nanningensis* (CP3) – **habitat type: probably lentic ('LeC')**

Paleo-Lake Nanning

Bellamyia cf. unicolor (CP4) – **habitat type: probably lotic ('LoA')**

Pickford (2004) suggested the Iriri Member of the Napak formation to be fluvio-paludal (i.e., slow-flowing or standing water) based on sedimentology, but interpreted the fossils as indicating a riverine environment rather than paludal conditions based on the presence of *Etheria*, the river oyster, which, however, can live in both fluvial and lacustrine conditions.

†*Neothauma hattinghi* (CP5) – **habitat type: uncertain ('LeB/LoB')**

Neothauma hattinghi was considered to have evolved in a large and probably shallow lake according to Van Damme and Pickford (1999). However, the habitat type was inferred based on the sculpture of the specimens only, and no other sedimentological evidence was provided. To avoid circular reasoning, we did not constrain the habitat type of this species.

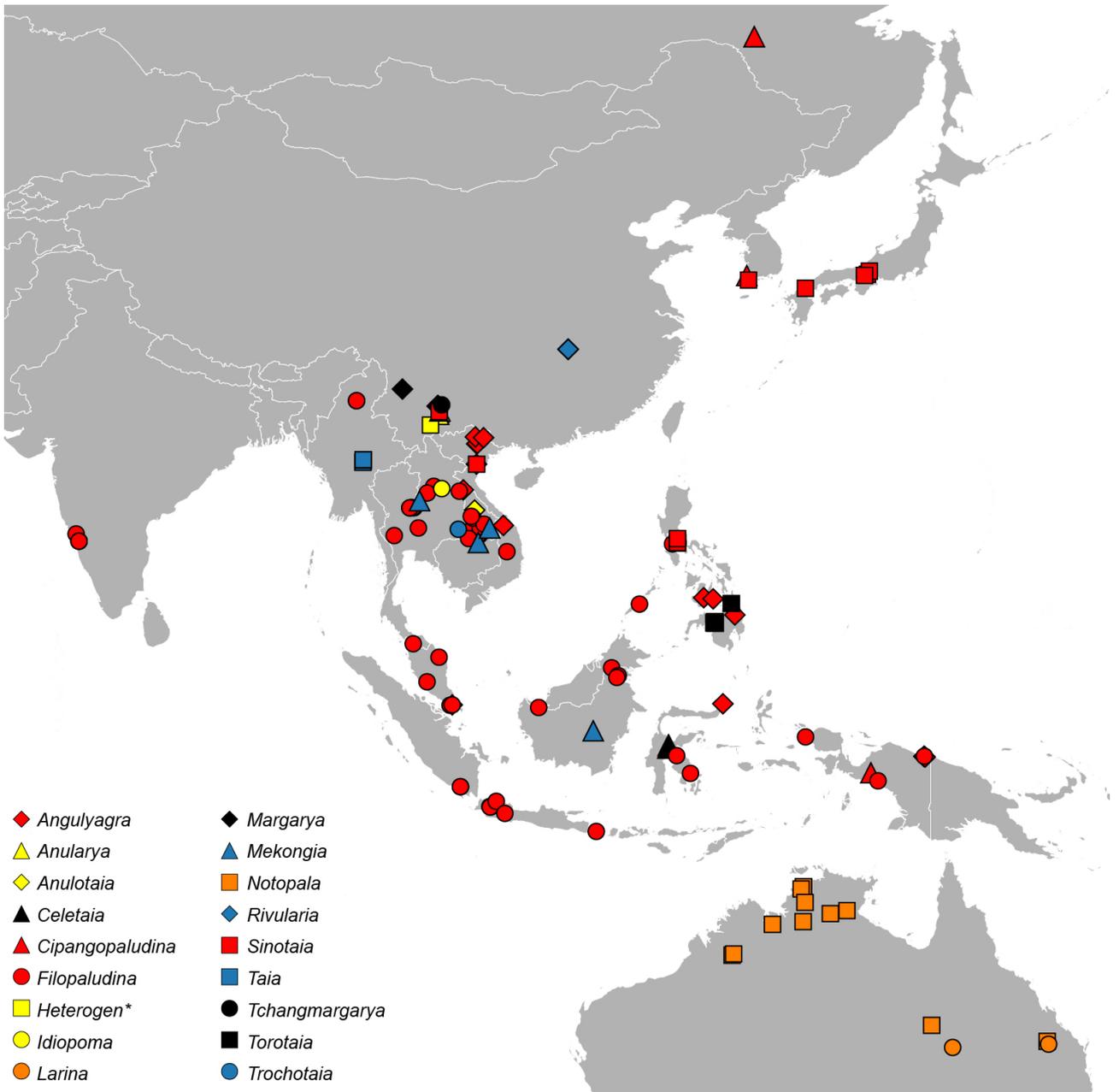


FIGURE S1. Distribution map of samples of Viviparidae collected in Asia/Australia. Note that a recent study has suggested to synonymize *Heterogen* with *Cipangopaludina* (Hirano et al. 2015).



FIGURE S2. Best Maximum Likelihood phylogram based on a RAXML analysis of the complete dataset (193 sequences). Numbers on branches indicate bootstrap values based on 456 replicates. See main text for details.

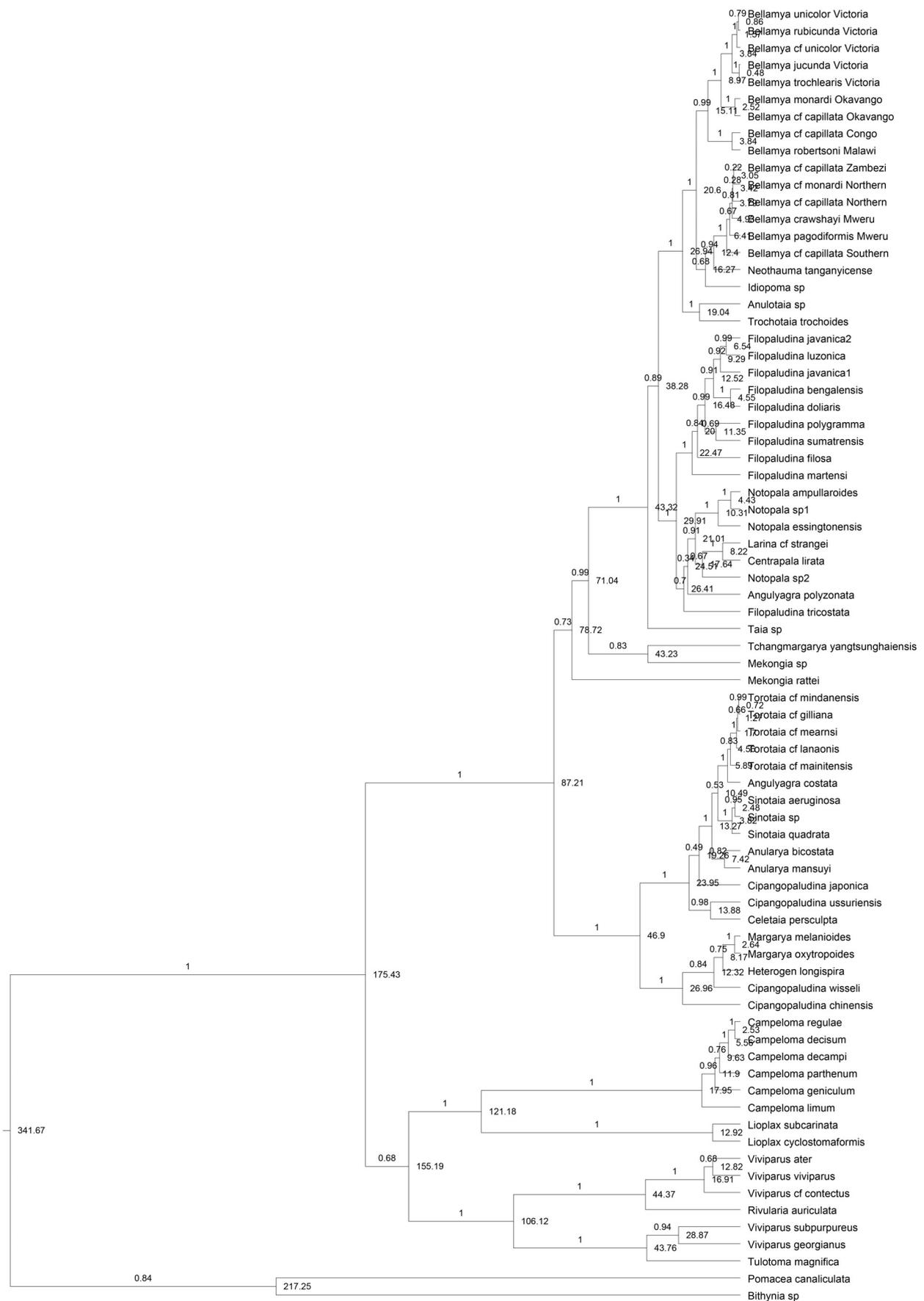


FIGURE S4. Maximum-clade credibility (MCC) tree based on a BEAST analysis of the reduced dataset (76 sequences) using five fossil-calibration points. Numbers on branches indicate posterior probabilities; numbers at nodes denote mean ages. See main text for details.

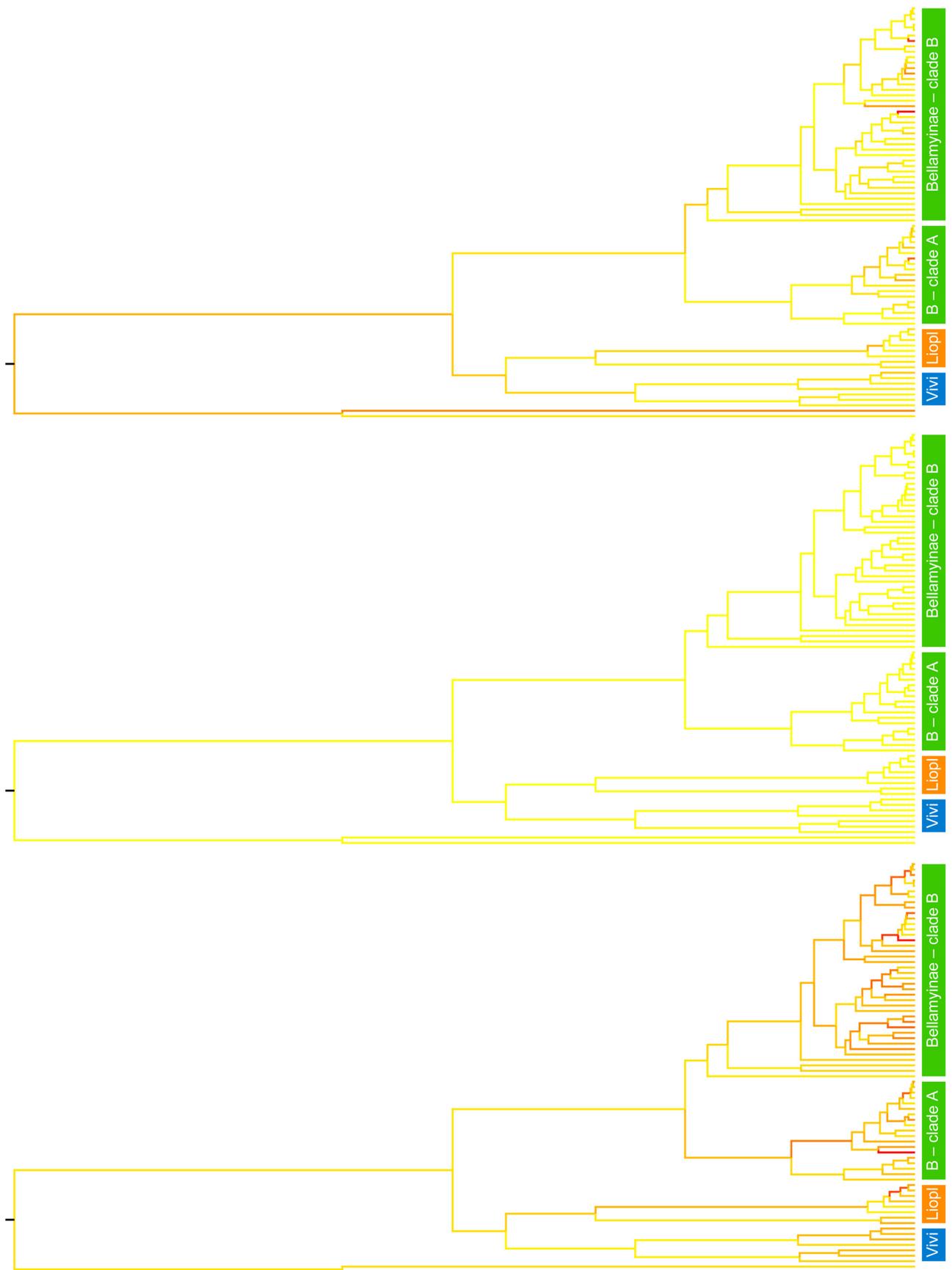


FIGURE S5. Branch-specific rates for nuclear 28S rRNA (upper panel), nuclear H3 (middle), and mitochondrial COI (lower panel) based on the BEAST MCC tree visualized in FigTree 1.4.2. The colour gradient across the branches denotes low (yellow) or high (red) mean rates. See Figure S4 for details.

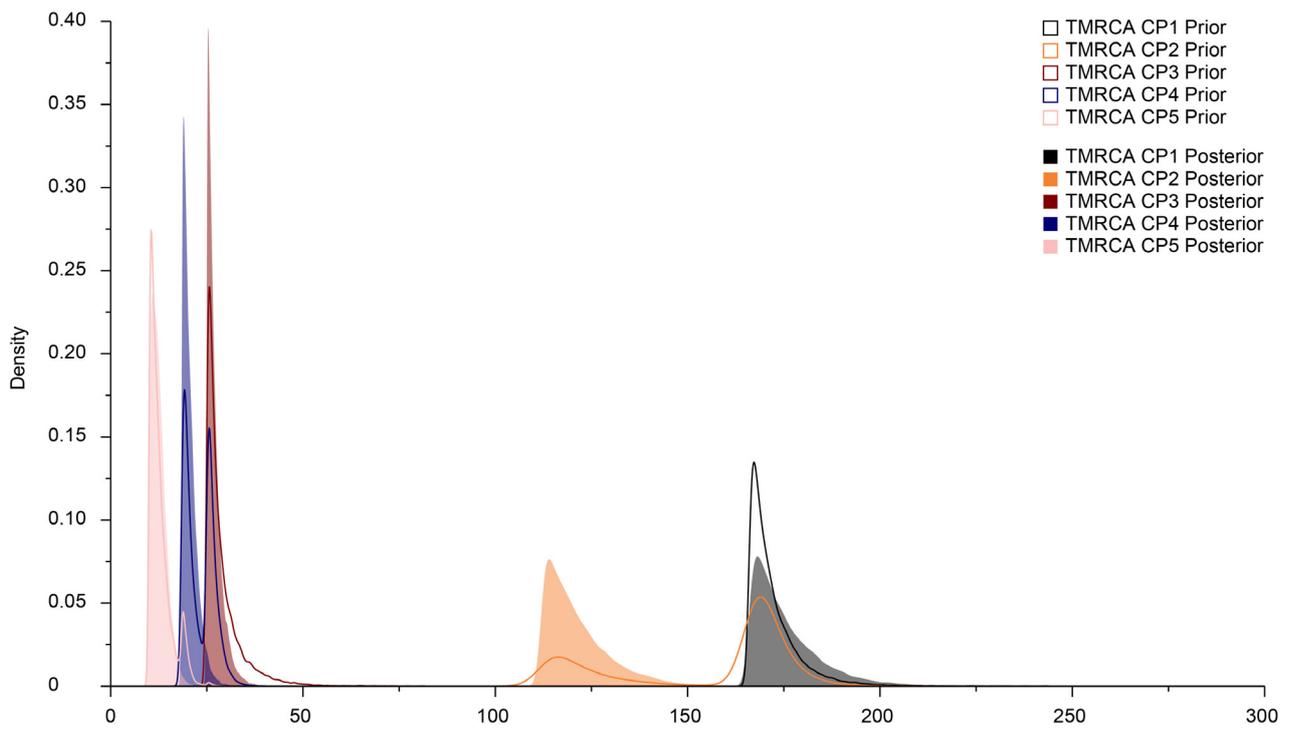


FIGURE S6. Comparison of posterior distributions obtained from the default BEAST analysis (using calibration points CP1-CP5) and the full prior distribution obtained the ‘sample from prior only’ analysis visualized in TRACER 1.5. See main text for details.



FIGURE S7. General shell morphology of Viviparidae. Voucher specimens of each representative operational taxonomic unit (OTU) shown in the OTU-based tree of Viviparidae (see Table S1). Additional image sources: *Anularya bicostata*, *Margarya oxytropoides* and *Tchangmargarya yangtsunghaiensis* from Zhang et al. (2015); *Bellamyia crawshayi* from Sengupta et al. (2009), *Cipangopaludina ussuriensis* from Gerstfeldt (1859); *Cipangopaludina japonica* from Van Bocxlaer and Strong (2016); and *Cipangopaludina longispira* from Hirano et al. (2015). *Bellamyia* species were assigned to biogeographical clades identified in the study of Schultheiß et al. (2014).

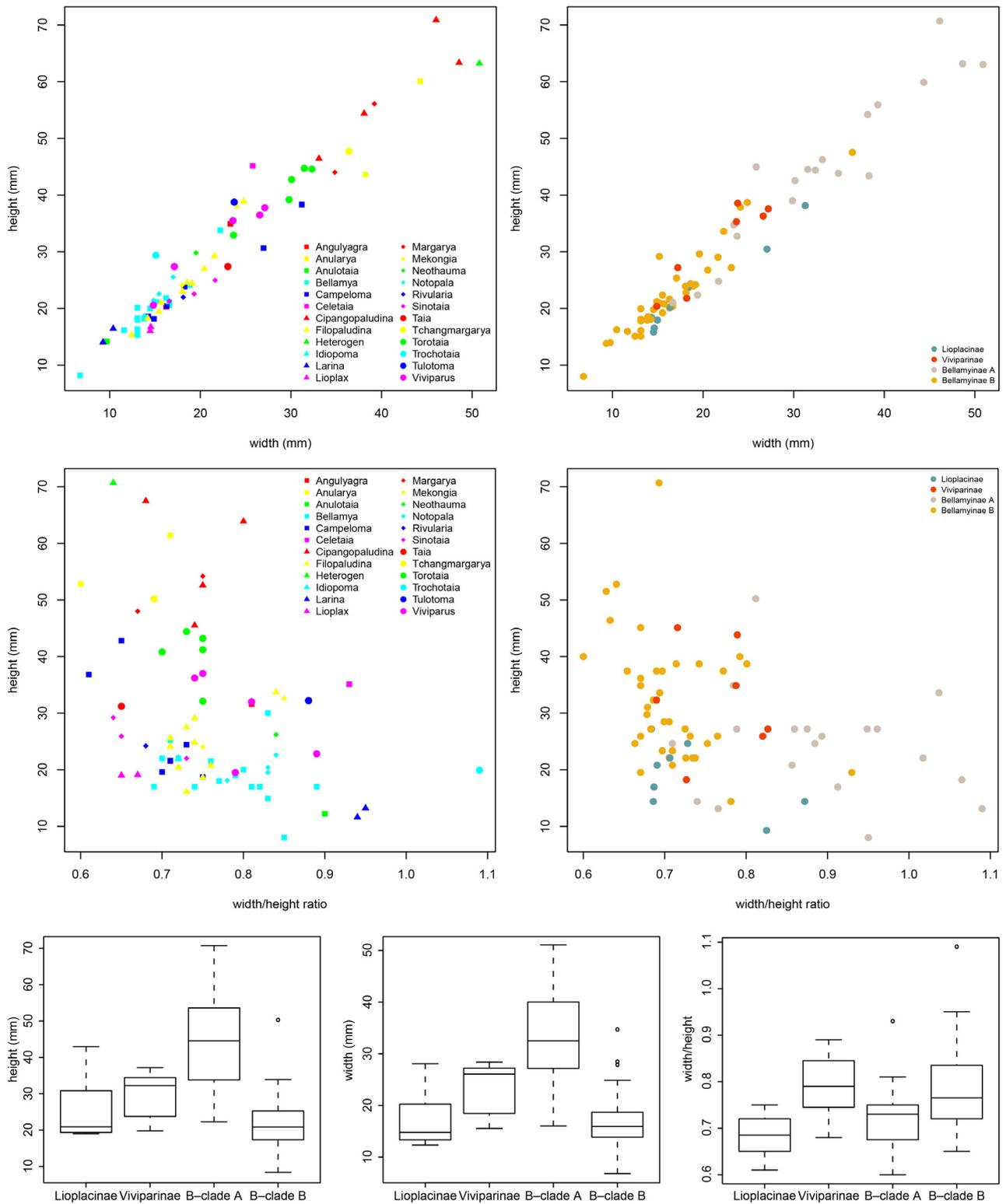


FIGURE S8. Genus- and clade-specific comparisons of standard shell measurements (height, width, and width-height ratio) obtained from the voucher images for each of the 74 OTUs shown in Fig. S6. Upper panels: width vs. height, middle panels: width-height ratio vs. height, and lower panels: boxplots per clade.

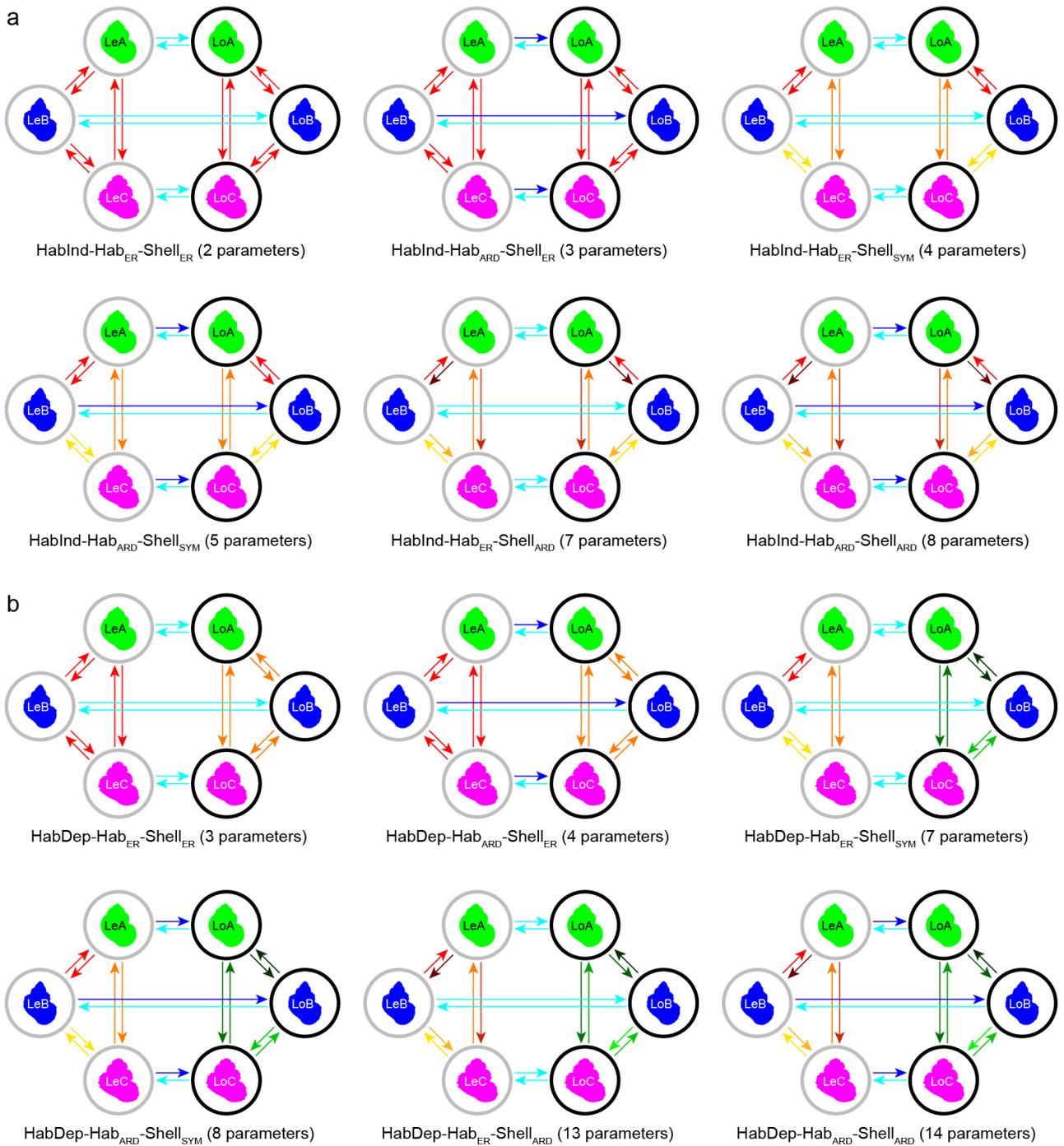


FIGURE S9. Illustration of transition matrices used to identify the best-fit model for habitat and shell transitions. a) Matrices for habitat-independent shell transitions; b) Matrices for habitat-dependent shell transitions. Note that transitions implementing both a habitat and shell shift (e.g., ‘LeA’→‘LoB’) were not allowed *a priori*. Character states: ‘LeA’ = lentic with shell type A (‘absent’), ‘LeB’ = lentic with shell type B (‘fine’), ‘LeC’ = lentic with shell type C (‘coarse’), ‘LoA’ = lotic with shell type A (‘absent’), ‘LoB’ = lotic with shell type B (‘fine’), and ‘LoC’ = lotic with shell type C (‘coarse’). Model specifications: ‘HabDep’ = habitat-dependent, ‘HabInd’ = habitat-independent, ‘Hab’ = habitat transition rate, ‘Shell’ = shell transition rate, ‘ARD’ = all rates different, ‘ER’ = equal rates, and ‘SYM’ = symmetric rates.

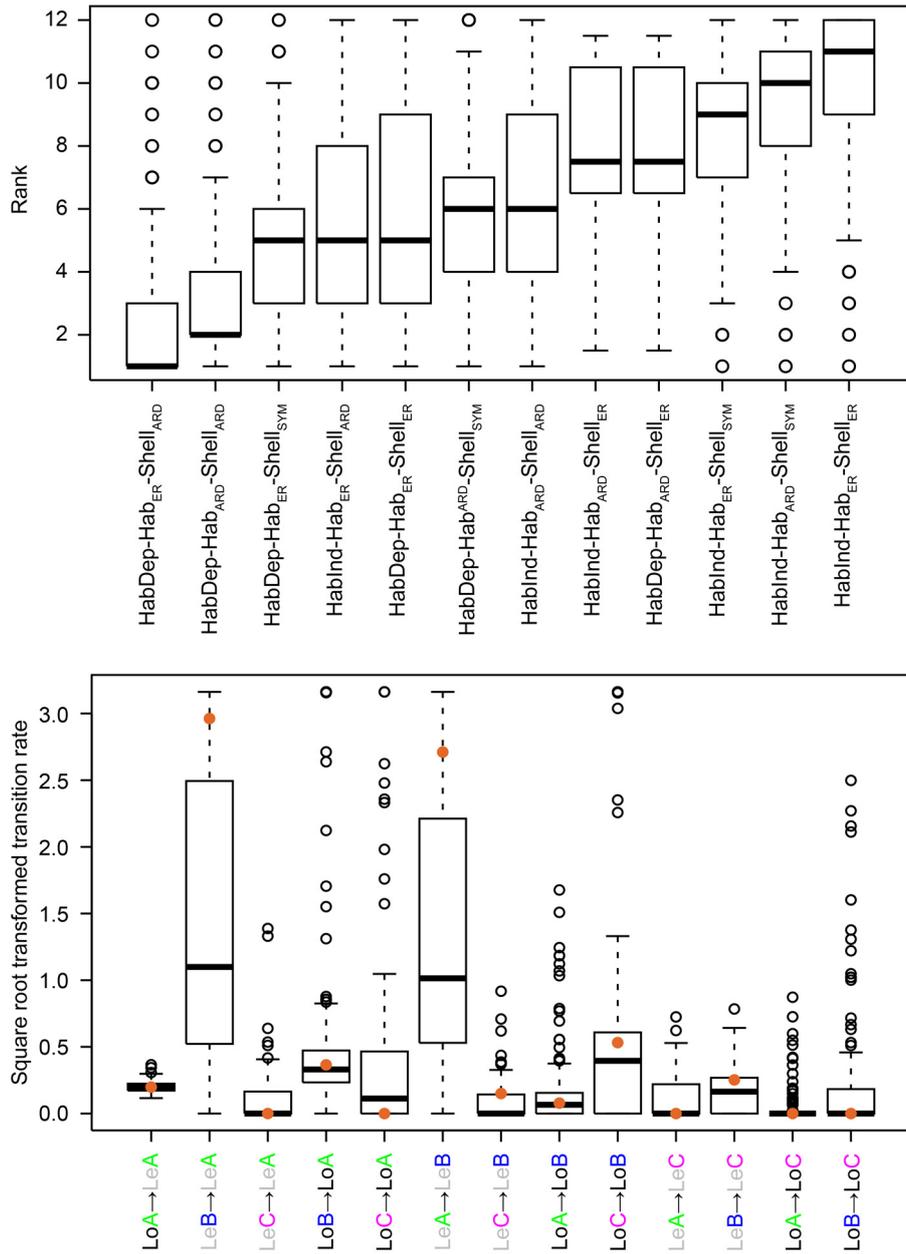


FIGURE S10. Evaluation of taxon sampling on model robustness based on 250 replicates (see main text for details). Upper panel: ranked model-fit comparison of habitat-shell evolution models according to AIC values (see also Fig. S9 and Table S8). Lower panel: estimated transition rates for the prevailing model of habitat-dependent asymmetric shell transition rates. Orange dots represent the transition rates estimated for the best-fit model in the unconstrained analysis that were used for simulations (see Table S8).

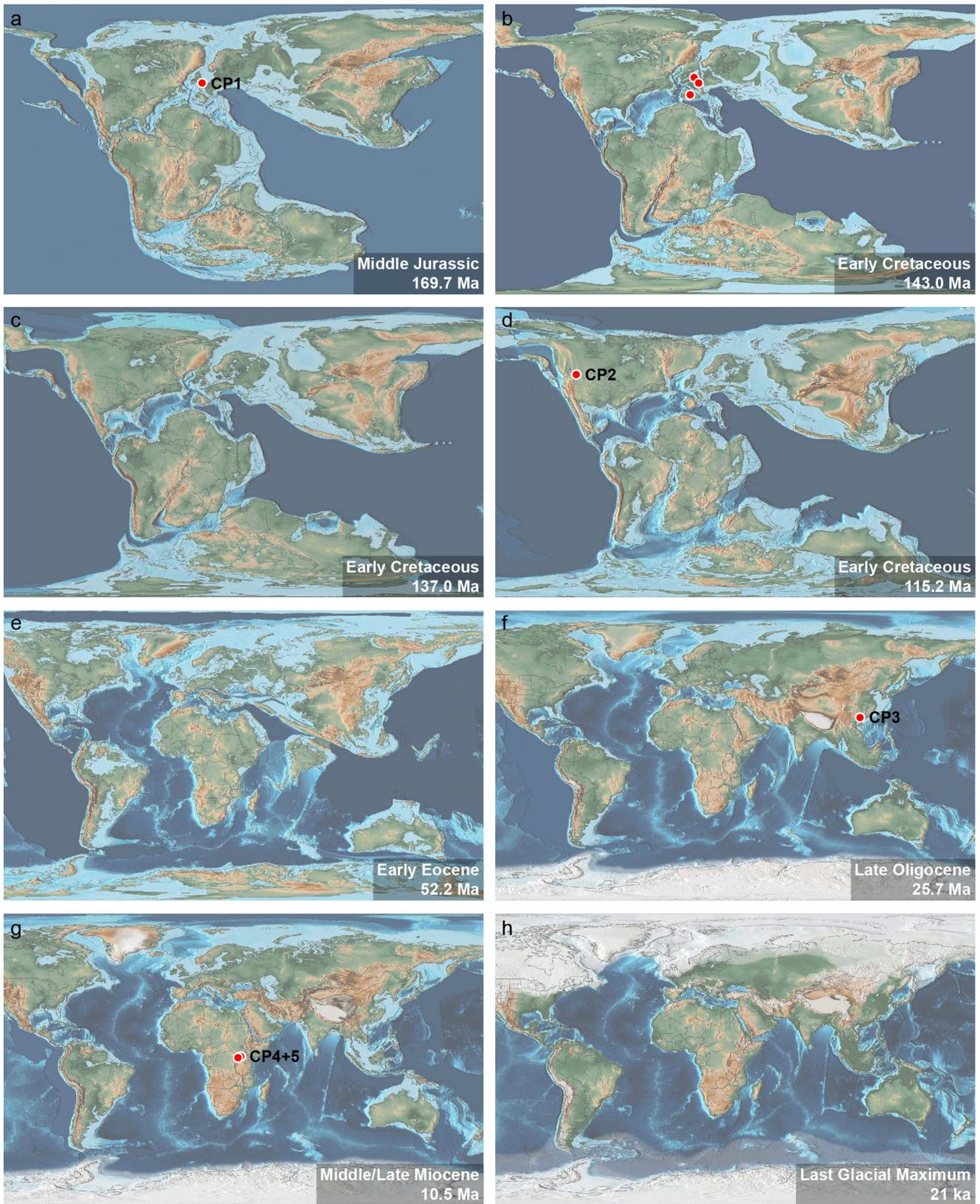


FIGURE S11. Palaeogeographical maps including selected fossil representatives mentioned in the main text. a) †*Viviparus langtonensis* (calibration point, CP1); b) †*Viviparus* spp.; c) Valanginian stage during which the colonization of Laurasia intensified; d) †*Campeloma harlowtonense* (CP2); e) onset of India-Asia collision; f) onset of Asia-Australia collision, †*Margarya nanningensis* (CP3); g) *Bellamyia* cf. *unicolor* (CP4, Early Miocene) and †*Neothauma hattinghi* (CP5); and h) the Last Glacial Maximum and the related global low sea level stand led to the interconnection of Southeast Asian islands. Modified from Scotese (2014a, 2014b, 2014c, 2014d).

TABLE S1. Material examined including locality information, assigned OTUs, predefined BIOGEOBEARS areas and GenBank accession numbers. *Bellamyia* species were assigned to biogeographical clades identified in the study of (Schultheiß et al. 2014). BIOGEOBEARS areas: (A) = North America, (B) = Africa, (C) = Europe, (D) = East Asia (China, Far East Russia, Japan, and South Korea), (E) = India, (F) = Indochina (incl. the Thai-Malay Peninsula and Singapore), (G) = Sumatra, (H) = Philippines, (J) Palawan, (K) = Borneo, (L) = Java (incl. Bali), (M) = Sulawesi, and (N) = ‘East of Wallacea’ (Australia and New Guinea). COI sequences marked with an asterisk were sequenced on a Roche 454 GS FLX Titanium platform (see main text and Table S2 for details). Specimens displayed in Fig. S7 are indicated with a camera icon.

Taxon/Specimen (incl. collection number)	OTU (BIOGEOBEARS)	BIOGEOBEARS area	Latitude	Longitude	COI	GenBank acc. no.	
						28S rRNA	H3
<i>Angulyagra costata</i> ZMB 107071-1 Sulawesi	<i>Angulyagra_costata</i>	H, M, N	1.49255	124.86733	MN997886	MN999555	MN997713
<i>Angulyagra costata</i> ZMB 113429-1 Cebu			9.79963	123.37553	MN997887*	MN999556	MN997714
<i>Angulyagra costata</i> ZMB 113431-1 Bohol			9.71163	124.10770	MN997888	MN999557	MN997715
<i>Angulyagra costata</i> ZMB 127100-1 LakeSentani			-2.62132	140.52718	MN997889	MN999558	MN997716
<i>Angulyagra costata</i> ZMB 127101-1 LakeSentani			-2.63322	140.49260	MN997890	MN999559	MN997717
<i>Angulyagra costata</i> ZMB 127109-1 LakeSentani			-2.68998	140.58362	MN997891	MN999560	MN997718
<i>Angulyagra costata</i> ZMB 230163-2 Mindanao			8.45128	125.79404	MH319893	MN999561	MN997719
<i>Angulyagra polyzonata</i> ZMB 114132-1 VNM	<i>Angulyagra_polyzonata</i>	F	20.25765	105.70833	MN997892	MN999562	MN997720
<i>Angulyagra polyzonata</i> ZMB 114171-1 VNM			21.84767	105.73150	MN997893	MN999563	MN997721
<i>Angulyagra polyzonata</i> ZMB 114175-1 VNM			21.88203	105.77450	MN997894	MN999564	MN997722
<i>Angulyagra polyzonata</i> ZMB 114189-1 VNM			22.40165	105.62417	MN997895	MN999565	MN997723
<i>Angulyagra polyzonata</i> ZMB 114202-1 VNM			22.33333	106.25833	MN997896	MN999566	MN997724
<i>Angulyagra polyzonata</i> ZMB 114403-1 VNM			15.46183	107.80183	MN997897	MN999567	MN997725
<i>Angulyagra polyzonata</i> ZMB 114411-1 LAO			18.24450	104.68683	MN997898	MN999568	MN997726
<i>Angulyagra polyzonata</i> ZMB 192077-1 SGP			1.40100	103.82150	MN997899	MN999569	MN997727
<i>Anularya bicostata</i> ZMB 193469-1 CHN	<i>Anularya_bicostata</i>	D	24.43150	102.86500	MN997900*	MN999570	MN997728
<i>Anularya mansuyi</i> ZMB 193466-3 CHN	<i>Anularya_mansuyi</i>	D	24.32967	102.76950	MN997901*	MN999571	MN997729
<i>Anularya mansuyi</i> ZMB 193466-5 CHN			24.32967	102.76950	MN997902	MN999572	MN997730
<i>Anularya mansuyi</i> ZMB 193467-1 CHN			24.16417	102.75883	MN997903*	MN999573	MN997731
<i>Anularya mansuyi</i> ZMB 193467-5 CHN			24.16417	102.75883	MN997904*	MN999574	MN997732
<i>Anulotaia</i> sp. ZMB 114963-1 LAO	<i>Anulotaia</i> sp	F	16.65800	105.56383	MN997905	MN999575	MN997733
<i>Bellamyia cf. capillata</i> 16985 ‘Congo’	<i>Bellamyia cf capillata</i> Congo	B	2.09981	22.66347	JX489246	JX489315	JX489284
<i>Bellamyia cf. capillata</i> 9473 ‘Northern’	<i>Bellamyia cf capillata</i> Northern	B	-9.04567	29.04460	HQ012725	HQ012692	JX489259
<i>Bellamyia cf. capillata</i> 17554 ‘Okavango’	<i>Bellamyia cf capillata</i> Okavango	B	-17.75630	25.18654	JX489255	JX489359	JX489293
<i>Bellamyia cf. capillata</i> 16566 ‘Southern’	<i>Bellamyia_cf_capillata_Southern</i>	B	-14.38287	23.23285	JX489243	JX489312	JX489279
<i>Bellamyia cf. capillata</i> 16567 ‘Southern’			-14.38287	23.23285	JX489244	JX489313	JX489280
<i>Bellamyia cf. capillata</i> 16569 ‘Southern’			-15.08967	22.84252	JX489245	JX489314	JX489281
<i>Bellamyia cf. capillata</i> 17539 ‘Southern’			-15.19352	22.95794	JX489251	JX489319	JX489289
<i>Bellamyia cf. capillata</i> 17544 ‘Southern’			-16.24204	23.24116	JX489252	JX489320	JX489290
<i>Bellamyia cf. capillata</i> 17545 ‘Southern’			-16.24204	23.24116	JX489253	JX489321	JX489291
<i>Bellamyia cf. capillata</i> 17549 ‘Southern’			-16.65075	23.56652	JX489254	JX489322	JX489292
<i>Bellamyia cf. capillata</i> 17053 ‘Zambezi’	<i>Bellamyia cf capillata</i> Zambezi	B	-13.55640	23.09447	JX489250	JX489318	JX489288

<i>Bellamyia cf. monardi</i> 16987 'Northern' 📷	Bellamyia_cf_monardi_Northern	B	-14.81798	26.04066	JX489248	JX489316	JX489286
<i>Bellamyia cf. monardi</i> 16995 'Northern'			-14.70344	26.10316	JX489249	JX489317	JX489287
<i>Bellamyia cf. unicolor</i> 15745 'Victoria' 📷	Bellamyia_cf_unicolor_Victoria	B	1.84843	30.47826	JX489240	JX489310	JX489277
<i>Bellamyia crawshayi</i> GB 'Mweru'	Bellamyia_crawshayi_Mweru	B	-8.99556	28.71611	FJ405844	FJ405695	FJ405746
<i>Bellamyia jucunda</i> 15321 'Victoria' 📷	Bellamyia_jucunda_Victoria	B	0.03892	32.49402	JX489231	JX489302	JX489269
<i>Bellamyia monardi</i> 9489 'Okavango' 📷	Bellamyia_monardi_Okavango	B	-18.12071	21.58754	HQ012800	HQ012686	JX489263
<i>Bellamyia pagodiformis</i> 9477 'Mweru' 📷	Bellamyia_pagodiformis_Mweru	B	-9.34801	28.73133	HQ012720	HQ012688	JX489261
<i>Bellamyia robertsoni</i> 6597 'Malawi' 📷	Bellamyia_robertsoni_Malawi	B	-14.08923	34.93017	HQ012797	HQ012704	JX489257
<i>Bellamyia rubicunda</i> 15312 'Victoria'	Bellamyia_rubicunda_Victoria	B	1.58649	31.10057	JX489230	JX489301	JX489268
<i>Bellamyia rubicunda</i> 15482 'Victoria' 📷			1.81842	31.32021	JX489233	JX489304	JX489271
<i>Bellamyia rubicunda</i> 15483 'Victoria'			1.81842	31.32021	JX489234	JX489305	JX489272
<i>Bellamyia trochlearis</i> 15192 'Victoria'	Bellamyia_trochlearis_Victoria	B	0.16176	33.58264	JX489227	JX489298	JX489265
<i>Bellamyia trochlearis</i> 15193 'Victoria'			0.16176	33.58264	JX489228	JX489299	JX489266
<i>Bellamyia trochlearis</i> 15494 'Victoria' 📷			0.03892	32.49402	JX489235	JX489339	JX489273
<i>Bellamyia unicolor</i> 15261 'Victoria' 📷	Bellamyia_unicolor_Victoria	B	0.14067	33.60258	JX489229	JX489300	JX489267
<i>Bellamyia unicolor</i> 15476 'Victoria'			2.45933	31.50487	JX489232	JX489303	JX489270
<i>Bellamyia unicolor</i> 15707 'Victoria'			1.09002	32.93648	JX489238	JX489307	JX489275
<i>Bellamyia unicolor</i> 15733 'Victoria'			1.69255	32.09473	JX489239	JX489309	JX489276
<i>Bithynia</i> sp. ZMB 191552-1 Sulawesi	Bithynia_sp	N.A. (outgroup)	-1.89580	120.66565	KY574006	N.A.	KY574014
<i>Campeloma decampi</i> USNM 1292572 USA 📷	Campeloma_decampi	A	34.68917	-86.82861	MN997906	MN999576	MN997734
<i>Campeloma decampi</i> USNM 1292573 USA		A	34.74278	-87.08444	MN997907	MN999577	MN997735
<i>Campeloma decisum</i> USNM 1292574 USA 📷	Campeloma_decisum	A	35.60389	-86.87528	MN997908	MN999578	MN997736
<i>Campeloma decisum</i> USNM 1292575 USA		A	34.75833	-87.81306	MN997909	MN999579	MN997737
<i>Campeloma geniculum</i> USNM 1292576 USA 📷	Campeloma_geniculum	A	31.00778	-87.16222	MN997910	MN999580	MN997738
<i>Campeloma limum</i> NCSM 27484 USA 📷	Campeloma_limum	A	32.44167	-81.83333	MN997911	MN999581	MN997739
<i>Campeloma parthenum</i> NCSM 45096 USA 📷	Campeloma_parthenum	A	31.42573	-88.45760	MN997912	MN999582	MN997740
<i>Campeloma regulae</i> USNM 1292577 USA	Campeloma_regulae	A	32.89639	-86.43472	MN997913	MN999583	MN997741
<i>Campeloma regulae</i> USNM 1292580 USA 📷		A	33.16806	-87.02139	MN997914	MN999584	MN997742
<i>Celetaia persculpta</i> ZMB 190992-1 Sulawesi	Celetaia_persculpta	M	-1.98372	120.58513	MN997915	MN999585	MN997743
<i>Celetaia persculpta</i> ZMB 190993-5 Sulawesi			-1.96547	120.67560	MN997916	MN999586	MN997744
<i>Celetaia persculpta</i> ZMB 191078-5 Sulawesi			-1.92128	120.67135	MN997917	MN999587	MN997745
<i>Celetaia persculpta</i> ZMB 192209a-1 Sulawesi			-1.82850	120.63593	MN997918*	MN999588	MN997746
<i>Celetaia persculpta</i> ZMB 192210a-1 Sulawesi			-1.74285	120.65202	MN997919	MN999589	MN997747
<i>Celetaia persculpta</i> ZMB 192211a-1 Sulawesi			-1.77105	120.62488	MN997920	MN999590	MN997748
<i>Celetaia persculpta</i> ZMB 192213a-1 Sulawesi 📷			-1.77323	120.63878	MN997921	MN999591	MN997749
<i>Cipangopaludina ussuriensis</i> UGSB 23018 RUS	Cipangopaludina_ussuriensis	D	53.73392	127.30203	MN997923	MN999593	MN997751
<i>Cipangopaludina ussuriensis</i> UGSN23019 RUS	Cipangopaludina_ussuriensis		53.75181	127.30603	MN997924	MN999594	MN997752
<i>Cipangopaludina chinensis</i> ZMB 192694-1 KOR 📷	Cipangopaludina_chinensis	D	35.03267	126.72100	MN997925*	MN999595	MN997753
<i>Cipangopaludina japonica</i> USNM 1296945 USA 📷	Cipangopaludina_japonica	D	38.81370	-77.03800	MN997926	MN999596	MN997754
<i>Cipangopaludina wisseli</i> ZMB 127112a LakePaniai	Cipangopaludina_wisseli	N	-3.91975	136.36817	MN997927	MN999597	MN997755
<i>Filopaludina bengalensis</i> ZMB 113662-2 IND	Filopaludina_bengalensis	E	14.79633	74.59083	MN997928	MN999598	MN997756

<i>Filopaludina bengalensis</i> ZMB 113664-1 IND			14.22833	74.81200	MN997929	MN999599	MN997757
<i>Filopaludina bengalensis</i> ZMB 113664-3 IND 📷			14.22833	74.81200	MN997930	MN999600	MN997758
<i>Filopaludina bengalensis</i> ZMB 113664-4 IND			14.22833	74.81200	MN997931	MN999601	MN997759
<i>Filopaludina decipiens</i> ZMB 127091-1 WestPapua	Filopaludina_javanica2	K, L, M, N	-1.10817	131.29255	MN997932	MN999602	MN997760
<i>Filopaludina decipiens</i> ZMB 127107-1 WestPapua 📷			-4.54932	136.92412	MN997933	MN999603	MN997761
<i>Filopaludina doliaris</i> ZMB 113665-4 MMR 📷	Filopaludina_doliaris	F	25.23383	96.39000	MN997934	MN999604	MN997762
<i>Filopaludina doliaris</i> ZMB 113665-5 MMR			25.23383	96.39000	MN997935	MN999605	MN997763
<i>Filopaludina filosa</i> ZMB 114012-1 THA 📷	Filopaludina_filosa	F	16.86687	100.63167	MN997936*	MN999606	MN997764
<i>Filopaludina filosa</i> ZMB 114049-1 THA			16.84375	100.85672	MN997937	MN999607	MN997765
<i>Filopaludina filosa</i> ZMB 192078-1 SPG			1.37198	103.66118	MN997938	MN999608	MN997766
<i>Filopaludina javanica</i> ZMB 112753-1 Sarawak	Filopaludina_javanica2	K, L, M, N	1.19733	110.54733	MN997939	MN999609	MN997767
<i>Filopaludina javanica</i> ZMB 127093-1 Kalimantan			4.30733	116.21750	MN997943	MN999613	MN997771
<i>Filopaludina javanica</i> ZMB 127096-1 Kalimantan			3.68967	116.73050	MN997944	MN999614	MN997772
<i>Filopaludina javanica</i> ZMB 127098-1 Kalimantan			3.55188	116.62385	MN997945	MN999615	MN997773
<i>Filopaludina javanica</i> ZMB 191097-1 Sulawesi			-2.57760	121.26892	MN997946	MN999616	MN997774
<i>Filopaludina javanica</i> ZMB 191406-1 Bali			-8.51243	115.02283	MN997948	MN999618	MN997776
<i>Filopaludina javanica</i> ZMB 127082-1 Java	Filopaludina_javanica1	G, L, M	-6.55548	106.74945	MN997940	MN999610	MN997768
<i>Filopaludina javanica</i> ZMB 127085-1 Java			-6.60083	106.79867	MN997941	MN999611	MN997769
<i>Filopaludina javanica</i> ZMB 127086-1 Java 📷			-6.59800	106.77773	MN997942	MN999612	MN997770
<i>Filopaludina javanica</i> ZMB 191398-3 Sulawesi			-3.96495	122.34565	MN997947	MN999617	MN997775
<i>Filopaludina javanica</i> ZMB 191702-2 Sumatra			-5.00882	104.46482	MN997949	MN999619	MN997777
<i>Filopaludina javanica</i> ZMB 191702-3 Sumatra			-5.00882	104.46482	MN997950	MN999620	MN997778
<i>Filopaludina javanica</i> ZMB 192080-1 Java			-6.15330	107.24835	MN997951	MN999621	MN997779
<i>Filopaludina luzonica</i> FLMNH 113451 Palawan	Filopaludina_luzonica	H, J	9.30000	118.38333	MN997952	MN999622	MN997780
<i>Filopaludina luzonica</i> ZMB 113414-1 Luzon			14.11340	121.36477	MN997953	MN999623	MN997781
<i>Filopaludina luzonica</i> ZMB 113417-1 Luzon			14.01517	120.95417	MN997954	MN999624	MN997782
<i>Filopaludina luzonica</i> ZMB 113437-1 Luzon			14.10620	121.37590	MN997955	MN999625	MN997783
<i>Filopaludina luzonica</i> ZMB 113438-1 Luzon			14.11730	121.36842	MN997956*	MN999626	MN997784
<i>Filopaludina luzonica</i> ZMB 113439-1 Luzon 📷			14.12242	121.33663	MN997957*	MN999627	MN997785
<i>Filopaludina martensi</i> ZMB 113166-1 LAO	Filopaludina_martensi	F	14.72450	105.95150	MN997958	MN999628	MN997786
<i>Filopaludina martensi</i> ZMB 113176-1 THA			14.95217	105.00917	MN997959*	MN999629	MN997787
<i>Filopaludina martensi</i> ZMB 113178-2 LAO			14.80833	105.92000	MN997960*	MN999630	MN997788
<i>Filopaludina martensi</i> ZMB 114000-1 THA			15.26433	101.19367	MN997961	MN999631	MN997789
<i>Filopaludina martensi</i> ZMB 114019-1 THA			16.83783	100.53017	MN997962*	MN999632	MN997790
<i>Filopaludina martensi</i> ZMB 114036-1 THA			16.84567	100.75027	MN997963	MN999633	MN997791
<i>Filopaludina martensi</i> ZMB 114120-1 THA			16.83967	100.53863	MN997964	MN999634	MN997792
<i>Filopaludina martensi</i> ZMB 114412-1 LAO			18.14417	104.37533	MN997965	MN999635	MN997793
<i>Filopaludina martensi</i> ZMB 114951-1 THA 📷			14.66750	99.31833	MN997966*	MN999636	MN997794
<i>Filopaludina martensi</i> ZMB 193437-1 LAO			18.52762	102.39233	MN997967	MN999637	MN997795
<i>Filopaludina polygramma</i> ZMB 113167-1 LAO	Filopaludina_polygramma	F	15.33533	105.98183	MN997968	MN999638	MN997796
<i>Filopaludina polygramma</i> ZMB 113170-1 LAO			16.03100	105.43867	MN997969	MN999639	MN997797
<i>Filopaludina polygramma</i> ZMB 114948-1 LAO 📷			15.56417	106.29150	MN997970	MN999640	MN997798

<i>Filopaludina polygramma</i> ZMB 114959-1 LAO			16.17433	105.30850	MN997971	MN999641	MN997799
<i>Filopaludina sumatrensis</i> ZMB 113175-1 THA	<i>Filopaludina_sumatrensis</i>	F, L	14.43517	105.10500	MN997972*	MN999642	MN997800
<i>Filopaludina sumatrensis</i> ZMB 113444-1 MYS			5.13745	102.80230	MN997973	MN999643	MN997801
<i>Filopaludina sumatrensis</i> ZMB 113446 MYS			3.21073	101.86743	MN997974	MN999644	MN997802
<i>Filopaludina sumatrensis</i> ZMB 113447b-1 MYS			6.18982	100.79600	MN997975	MN999645	MN997803
<i>Filopaludina sumatrensis</i> ZMB 114070-1 THA 📷			18.00122	101.88633	MN997976*	MN999646	MN997804
<i>Filopaludina sumatrensis</i> ZMB 114405 VNM			13.40900	108.07483	MN997977	MN999647	MN997805
<i>Filopaludina sumatrensis</i> ZMB 191654 Java			-7.10020	107.91727	MN997978	MN999648	MN997806
<i>Filopaludina sumatrensis</i> ZMB 192076 SGP			1.40100	103.82150	MN997979	MN999649	MN997807
<i>Filopaludina tricostrata</i> ZMB 127114-1 LakeSentani 📷	<i>Filopaludina_tricostrata</i>	N	-2.62132	140.52718	MN997980	MN999650	MN997808
<i>Heterogen longispira</i> GB CHN	<i>Heterogen_longispira</i>	D	23.67000	102.57000	GU198809	KJ867091	N.A.
<i>Idiopoma</i> sp. ZMB 113172-1 LAO 📷	<i>Idiopoma_sp</i>	F	18.34667	103.00933	MN997981	MN999651	MN997809
<i>Idiopoma</i> sp. ZMB 192469-1 MMR	<i>Idiopoma_sp</i>		N.A.	N.A.	MN997982	MN999652	MN997810
<i>Larina cf. strangei</i> ZMB 192393 AUS 📷	<i>Larina cf strangei</i>	N	-25.18025	150.18370	MN997983	MN999653	MN997811
<i>Larina lirata</i> ZMB 192040 AUS 📷	<i>Larina lirata</i>	N	-25.43667	142.72700	MN997922	MN999592	MN997750
<i>Lioplax cyclostomaformis</i> USNM 1292582 USA 📷	<i>Lioplax_cyclostomaformis</i>	A	33.16806	-87.02139	MN997984	MN999654	MN997812
<i>Lioplax subcarinata</i> NCSM 40332-1 USA 📷z	<i>Lioplax_subcarinata</i>	A	35.81510	-77.54652	MN997985	MN999655	MN997813
<i>Lioplax subcarinata</i> USNM 1292583 USA 📷		A	34.75833	-87.81306	N.A.	MN999656	MN997814
<i>Margarya melanioides</i> ZMB 193463-2 CHN	<i>Margarya_melanioides</i>	D	24.80050	102.67133	MN997986*	MN999657	MN997815
<i>Margarya melanioides</i> ZMB 193463-7 CHN 📷			24.80050	102.67133	MN997987	MN999658	MN997816
<i>Margarya melanioides</i> ZMB 193463-8 CHN			24.80050	102.67133	MN997988*	MN999659	MN997817
<i>Margarya melanioides</i> ZMB 193468-1 CHN			26.14683	99.95250	MN997989	MN999660	MN997818
<i>Margarya oxytropoides</i> GB CHN	<i>Margarya_oxytropoides</i>	D	24.86694	102.71332	KJ636775	KJ867094	N.A.
<i>Mekongia rattei</i> ZMB 114062-1 THA	<i>Mekongia_rattei</i>	F	17.39268	101.28417	MN997990	MN999661	MN997819
<i>Mekongia rattei</i> ZMB 114956-1 LAO 📷			14.11667	105.85500	MN997991*	MN999662	MN997820
<i>Mekongia rattei</i> ZMB 114958-1 LAO			15.25600	106.73550	MN997992*	MN999663	MN997821
<i>Mekongia</i> sp. ZMB 192085-1 Kalimantan 📷	<i>Mekongia_sp</i>	K	-0.63083	114.78783	MN997993	MN999664	MN997822
<i>Neothauma tanganyicense</i> 6992 7033 Tanganyika 📷	<i>Neothauma_tanganyicense</i>	B	-4.78543	29.69946	HQ012717	HQ012707	MN997823
<i>Neothauma tanganyicense</i> ZMB 107101-1 Tanganyika			-8.72467	31.13300	MN997994	MN999665	MN997824
<i>Notopala ampullaroides</i> ZMB 106669-1 AUS 📷	<i>Notopala_ampullaroides</i>	N	-14.07067	131.25093	MN997995	MN999666	MN997825
<i>Notopala ampullaroides</i> ZMB 106691-1 AUS			-15.79215	128.71755	MN997996	MN999667	MN997826
<i>Notopala essingtonensis</i> ZMB 106602-1 AUS	<i>Notopala_essingtonensis</i>	N	-12.83803	131.13330	MN997997	MN999668	MN997827
<i>Notopala essingtonensis</i> ZMB 106638-1 AUS 📷			-14.71337	134.50790	MN997998	MN999669	MN997828
<i>Notopala essingtonensis</i> ZMB 106647-1 AUS			-13.02193	130.95155	MN997999	MN999670	MN997829
<i>Notopala essingtonensis</i> ZMB 192707-1 AUS			-14.95263	133.22150	MN998000	MN999671	MN997830
<i>Notopala</i> sp. ZMB 106620-1 AUS 📷	<i>Notopala_sp1</i>	N	-15.58103	131.10237	MN998001	MN999672	MN997831
<i>Notopala</i> sp. ZMB 106694-1 AUS			-18.21088	125.57900	MN998002	MN999673	MN997832
<i>Notopala</i> sp. ZMB 106695-1 AUS			-18.10868	125.69818	MN998003	MN999674	MN997833
<i>Notopala</i> sp. ZMB 192042-1 AUS	<i>Notopala_sp2</i>	N	-23.70367	141.09583	MN998004	MN999675	MN997834
<i>Notopala</i> sp. ZMB 192392-1 AUS 📷			-24.95695	150.07405	MN998005	MN999676	MN997835
<i>Pomacea canaliculata</i> ZMB 191395a Sulawesi	<i>Pomacea_canaliculata</i>	N.A. (outgroup)	-5.35915	119.47758	KY574007	KY573999	KY574015

<i>Rivularia auriculata</i> ZMB 116176-1 CHN	<i>Rivularia_auriculata</i>	D	29.26350	112.83800	KY574008	KY574000	KY574016
<i>Rivularia auriculata</i> ZMB 116176-2 CHN 📷			29.26350	112.83800	KY574009	KY574001	KY574017
<i>Rivularia auriculata</i> ZMB 116176-3 CHN			29.26350	112.83800	KY574010	KY574002	KY574018
<i>Sinotaia aeruginosa</i> ZMB 114131-1 VNM 📷	<i>Sinotaia_aeruginosa</i>	F	20.25765	105.72500	MN998006	MN999677	MN997836
<i>Sinotaia quadrata</i> ZMB 113413-1 Luzon 📷	<i>Sinotaia_quadrata</i>	D, H	14.07825	121.34230	MN998007	MN999678	MN997837
<i>Sinotaia quadrata</i> ZMB 113416-1 Luzon			14.11377	121.33817	MN998008*	MN999679	MN997838
<i>Sinotaia quadrata</i> ZMB 113442-1 Luzon			14.42767	121.33700	MN998009*	MN999680	MN997839
<i>Sinotaia quadrata</i> ZMB 192692-1 JPN			35.13542	136.09883	MN998010	MN999681	MN997840
<i>Sinotaia quadrata</i> ZMB 192730-1 JPN			35.38333	136.25333	MN998011	MN999682	MN997841
<i>Sinotaia quadrata</i> ZMB 192731-1 JPN			34.04167	131.28333	MN998012	MN999683	MN997842
<i>Sinotaia quadrata</i> ZMB 192732-1 KOR			34.68567	126.86167	MN998013	MN999684	MN997843
<i>Sinotaia quadrata</i> ZMB 192735-1 JPN			35.05333	135.87500	MN998014*	MN999685	MN997844
<i>Sinotaia</i> sp. ZMB 193457-2 CHN	<i>Sinotaia_sp</i>	D	24.37622	102.81633	MN998015	MN999686	MN997845
<i>Sinotaia</i> sp. ZMB 193457-5 CHN 📷			24.37622	102.81633	MN998016*	MN999687	MN997846
<i>Taia</i> sp. ZMB 113650-1 MMR	<i>Taia_sp</i>	F	20.55900	96.90300	MN998017	MN999688	MN997847
<i>Taia</i> sp. ZMB 113654-4 MMR			20.42100	96.88667	MN998018	MN999689	MN997848
<i>Taia</i> sp. ZMB 113654-7 MMR 📷			20.42100	96.88667	MN998019	MN999690	MN997849
<i>Taia</i> sp. ZMB 113660-4 MMR			20.60750	96.90350	MN998020	MN999691	MN997850
<i>Taia</i> sp. ZMB 113660-7 MMR			20.60750	96.90350	MN998021	MN999692	MN997851
<i>Taia</i> sp. ZMB 113660-8 MMR			20.60750	96.90350	MN998022	MN999693	MN997852
<i>Taia</i> sp. ZMB 113660-9 MMR			20.60750	96.90350	MN998023	MN999694	MN997853
<i>Tchangmargarya yangtsunghaiensis</i> GB CHN	<i>Tchangmargarya_yangtsunghaiensis</i>	D	24.90944	103.00444	KJ636773	KJ867100	N.A.
<i>Torotaia</i> cf. <i>gilliana</i> UGSB 18385 Lanao 📷	<i>Torotaia_cf_gilliana</i>	H	7.87518	124.18552	MH319885	MN999695	MN997854
<i>Torotaia</i> cf. <i>lanaonis</i> UGSB 18372 Lanao	<i>Torotaia_cf_lanaonis</i>	H	7.80100	124.19423	MH319881	MN999696	MN997855
<i>Torotaia</i> cf. <i>lanaonis</i> UGSB 18377 Lanao			7.80100	124.19423	MH319883	MN999697	MN997856
<i>Torotaia</i> cf. <i>lanaonis</i> UGSB 18387 Lanao 📷			7.87518	124.18552	MH319886	MN999698	MN997857
<i>Torotaia</i> cf. <i>lanaonis</i> UGSB 18399 Lanao			7.79040	124.27493	MH319891	MN999699	MN997858
<i>Torotaia</i> cf. <i>mainitensis</i> ZMB 230165-1 Mindanao	<i>Torotaia_cf_mainitensis</i>	H	9.34755	125.52062	MH319896	MN999700	MN997859
<i>Torotaia</i> cf. <i>mearnsi</i> UGSB 18390 Lanao	<i>Torotaia_cf_mearnsi</i>	H	7.93712	124.23564	MH319887	MN999701	MN997860
<i>Torotaia</i> cf. <i>mearnsi</i> UGSB 18391 Lanao 📷			7.93712	124.23564	MH319888	MN999702	MN997861
<i>Torotaia</i> cf. <i>mindanensis</i> UGSB 18380 Lanao 📷	<i>Torotaia_cf_mindanensis</i>	H	7.83259	124.15815	MH319884	MN999703	MN997862
<i>Trochotaia trochoides</i> ZMB 113179-1 THA 📷	<i>Trochotaia_trochoides</i>	F	15.16100	104.29950	MN998024	MN999704	MN997863
<i>Tulotoma magnifica</i> USNM 1292584 USA	<i>Tulotoma_magnifica</i>	A	32.86556	-86.32556	MN998025	MN999705	MN997864
<i>Tulotoma magnifica</i> USNM 1292586 USA 📷		A	33.42889	-86.35278	MN998026	MN999706	MN997865
<i>Viviparus ater</i> GB CHE	<i>Viviparus_ater</i>	C	47.31667	8.58333	FJ405882	FJ405630	FJ405774
<i>Viviparus</i> cf. <i>contectus</i> GB MKD	<i>Viviparus_cf_contectus</i>	C	41.10222	20.80222	KY574011	KY574003	KY574019
<i>Viviparus georgianus</i> USNM 1292587 USA 📷	<i>Viviparus_georgianus</i>	A	34.07306	-87.07306	MN998027	MN999707	MN997866
<i>Viviparus subpurpureus</i> USNM 1292588 USA 📷	<i>Viviparus_subpurpureus</i>	A	34.75833	-87.81306	N.A.	MN999708	MN997867
<i>Viviparus viviparus</i> GB DEU	<i>Viviparus_viviparus</i>	C	52.84167	14.11583	KY574013	KY574005	KY574021

TABLE S2. Main sequencing information for all COI sequences that were obtained by amplicon sequencing on a Roche 454 GS FLX Titanium platform as indicated with an asterisk in Table S1 (see main text for details).

Taxon/Specimen (incl. collection number)	Number of reads	Coverage
<i>Angulyagra costata</i> ZMB113429-1 Cebu	55	15-55
<i>Anularya bicostata</i> ZMB193469-1 CHN	54	7-51
<i>Anularya mansuyi</i> ZMB193466-3 CHN	68	15-65
<i>Anularya mansuyi</i> ZMB193467-1 CHN	14	4-14
<i>Anularya mansuyi</i> ZMB193467-5 CHN	19	6-19
<i>Celetaia persculpta</i> ZMB192209a-1 Sulawesi	81	37-79
<i>Cipangopaludina chinensis</i> ZMB192694-1 KOR	174	35-75
<i>Filopaludina filosa</i> ZMB114012-1 THA	27	4-23
<i>Filopaludina luzonica</i> ZMB113438-1 Luzon	95	14-62
<i>Filopaludina luzonica</i> ZMB113439-1 Luzon	120	15-94
<i>Filopaludina martensi</i> ZMB113176-1 THA	14	5-13
<i>Filopaludina martensi</i> ZMB113178-2 LAO	36	16-34
<i>Filopaludina martensi</i> ZMB114019-1 THA	123	5-112
<i>Filopaludina martensi</i> ZMB114951-1 THA	33	6-32
<i>Filopaludina sumatrensis</i> ZMB113175-1 THA	23	7-22
<i>Filopaludina sumatrensis</i> ZMB114070-1 THA	17	1-17
<i>Margarya melanioides</i> ZMB193463-2 CHN	22	6-22
<i>Margarya melanioides</i> ZMB193463-8 CHN	16	5-16
<i>Mekongia rattei</i> ZMB114956-1 LAO	44	12-43
<i>Mekongia rattei</i> ZMB114958-1 LAO	25	5-24
<i>Sinotaia quadrata</i> ZMB113416-1 Luzon	52	6-51
<i>Sinotaia quadrata</i> ZMB113442-1 Luzon	66	7-58
<i>Sinotaia quadrata</i> ZMB192735-1 JPN	16	4-15
<i>Sinotaia</i> sp. ZMB193457-5 CHN	118	27-111

TABLE S3. Fossils used to calibrate the molecular phylogeny (see also ‘Brief Discussion of Fossils not Used for Calibration’ above).

Calibration point	Fossil species	Locality Formation	Stratigraphic age (Myr)	Reference
CP1	† <i>Viviparus langtonensis</i>	England, UK Inferior Oolite (Middle Jurassic, Bathonian)	174-166	Hudleston (1896); Tracey et al. (1993)
CP2	† <i>Campeloma harlowtonense</i>	Montana, USA Kootenai Formation (Early Cretaceous, Aptian)	121-112	Stanton (1903); Yen (1950)
CP3	† <i>Margarya nanningensis</i>	Guangxi, China Yongning Formation (Early? Oligocene)	30-25	Quan et al. (2016); Tian et al. (2018)
CP4	<i>Bellamya</i> cf. <i>unicolor</i>	Napak, Uganda Iriiri Member (Early Miocene)	20-18.5	Pickford (2004)
CP5	† <i>Neothauma hattinghi</i>	Albertine Rift Valley, Uganda Kakara and Lower Oluka Formation (Middle/Late Miocene)	11-10	Van Damme and Pickford (1999)

TABLE S4. Synapomorphic shell features for the fossils used to calibrate the molecular phylogeny.

Calibration point	Fossil species	Synapomorphic shell features	Reference
CP1	† <i>Viviparus langtonensis</i>	Shells of Viviparidae possess a high shell spire and a thick periostracum and thus differ from those of Ampullariidae and Cyclophoridae (all belonging to the Architaenioglossa)	Simone (2004)
CP2	† <i>Campeloma harlowtonense</i>	Shells of Lioplacinae are thick and turreted with an outer lip being subangulated, sinuous or incurved at the base. In <i>Campeloma</i> , the outer lip of the aperture is nearly straight in lateral profile (as the fossil), whereas it is strongly sinuous in <i>Lioplax</i>	Gill (1863); Thompson (1999)
CP3	† <i>Margarya nanningensis</i>	Shells of <i>Margarya</i> have a produced, ‘ <i>Melania</i> ’-like spire, are composed of scalariform and rapidly increasing whorls and are sculptured with prominent spiral ribs (keels)	Nevill (1877)
CP4	<i>Bellamya</i> cf. <i>unicolor</i>	Protoconchs of <i>Bellamya</i> are pointed and carinated	Van Damme and Pickford (1999)
CP5	† <i>Neothauma hattinghi</i>	Protoconchs of <i>Neothauma</i> are smooth and obtuse	Van Damme and Pickford (1999)

TABLE S5. Comparison of divergence time estimates for the nodes corresponding to the five fossil calibration points obtained from the original and modified BEAST analyses using a reduced set of CPs. Grey-marked divergence times refer to nodes that were not fossil-constrained.

Fossil calibration point	Full set of CPs (CP1-CP5)	Reduced set of CPs (CP1 only)	Reduced set of CPs (CP3-CP5 only)
CP1 mean (95% HPD)	175.4 (166.0, 192.9) My	174.9 (166.0, 192.8)	155.6 (84.0, 239.2) My
CP2 mean (95% HPD)	121.2 (112.0, 137.9) My	101.1 (53.7, 147.2) My	99.2 (49.2, 164.2) My
CP3 mean (95% HPD)	27.0 (25.0, 31.0) My	8.9 (3.0, 16.7) My	26.9 (25.0, 30.7) My
CP4 mean (95% HPD)	20.6 (18.5, 24.3) My	24.8 (12.3, 40.0) My	20.4 (18.5, 24.0) My
CP5 mean (95% HPD)	12.4 (10.0, 15.7) My	12.1 (4.3, 21.2) My	12.3 (10.0, 15.4) My

TABLE S6. BIOGEOBEARS results table. Best-fit models are printed in bold. ‘LnL’ = log likelihood, ‘Nparams’ = number of parameters, ‘d’ = dispersal rate, ‘e’ = extinction rate (i.e., range contraction), ‘j’ = jump dispersal weights, ‘AIC’ = Akaike information criterion.

Analysis	Model	LnL	Nparams	d	e	j	AIC	ΔAIC
unconstrained	DEC+J	-141.0	3	0.0005	0	0.0071	288.0	0
	DIVALIKE+J	-142.2	3	0.0006	0	0.0081	290.4	2.4
	BayAreaLIKE+J	-146.8	3	0.0004	0	0.0122	299.6	11.6
	DEC	-148.0	2	0.0009	0.0008	0	300.0	12.0
	DIVALIKE	-151.3	2	0.0010	0	0	306.6	18.6
	BayAreaLIKE	-181.5	2	0.0008	0.0099	0	367.0	79.0
fossil-constrained	DEC+J	-151.8	3	0.0006	0	0.0097	309.6	0
	DIVALIKE+J	-152.8	3	0.0006	0	0.0098	311.6	2.0
	BayAreaLIKE+J	-153.3	3	0.0004	0	0.0138	312.6	3.0
	DIVALIKE	-162.8	2	0.0012	0.0015	0	329.6	20.0
	DEC	-172.5	2	0.0020	0.0110	0	349.0	39.4
	BayAreaLIKE	-194.9	2	0.0017	0.0114	0	393.8	84.2

TABLE S7. Standard shell parameters for each voucher specimen shown in Fig. S7 as well as shell and habitat type assigned to each OTU and fossil species used to time-calibrate the phylogeny. Shell types: A = ‘sculpture absent’, B = ‘fine’, C = ‘coarse’ including noded spirals. Habitat type: ‘Le’ = lentic and ‘Lo’ = lotic.

Species (OTU)	Height (mm)	Width (mm)	Width/height	Shell type	Habitat type
<i>Angulyagra_costata</i>	31.6	25.6	0.81	B	Le/Lo ¹
<i>Angulyagra_polyzonata</i>	21.5	15.3	0.71	B	Le/Lo ¹
<i>Anularya_bicostata</i>	61.4	43.3	0.71	C	Le
<i>Anularya_mansuyi</i>	52.8	31.7	0.60	C	Le
<i>Anulotaia_sp</i>	12.2	11.0	0.90	B	Lo
<i>Bellamya_cf_capillata_Congo</i>	14.9	12.4	0.83	A	Lo
<i>Bellamya_cf_capillata_Northern</i>	25.2	18.0	0.71	A	Lo
<i>Bellamya_cf_capillata_Okavango</i>	17.0	13.7	0.81	A	Lo
<i>Bellamya_cf_capillata_Southern</i>	17.0	13.9	0.82	A	Lo
<i>Bellamya_cf_capillata_Zambezi</i>	18.0	13.8	0.77	A	Lo
<i>Bellamya_cf_monardi_Northern</i>	22.0	15.9	0.72	A	Lo
<i>Bellamya_cf_unicolor_Victoria</i>	8.0	6.8	0.85	A	Le
<i>Bellamya_crawshayi_Mweru</i>	21.5	16.4	0.76	B	Le
<i>Bellamya_jucunda_Victoria</i>	22.0	15.5	0.70	A	Le
<i>Bellamya_monardi_Okavango</i>	17.0	15.2	0.89	A	Lo
<i>Bellamya_pagodiformis_Mweru</i>	20.0	15.9	0.80	B	Le
<i>Bellamya_robertsoni_Malawi</i>	30.0	24.8	0.83	A	Le
<i>Bellamya_rubicunda_Victoria</i>	19.0	15.1	0.79	A	Le
<i>Bellamya_trochlearis_Victoria</i>	17.0	11.8	0.69	B	Le
<i>Bellamya_unicolor_Victoria</i>	17.0	12.5	0.74	A	Le
<i>Campeloma_decampi</i>	36.8	22.6	0.61	A	Lo
<i>Campeloma_decisum</i>	21.6	15.4	0.71	A	Lo
<i>Campeloma_geniculum</i>	18.7	14.1	0.75	A	Lo
<i>Campeloma_limum</i>	24.4	17.8	0.73	A	Lo
<i>Campeloma_parthenum</i>	42.8	28.0	0.65	A	Lo
<i>Campeloma_regulae</i>	19.6	13.8	0.70	A	Lo
<i>Celetaia_persculpta</i>	35.1	32.8	0.93	C	Le
<i>Cipangopaludina_ussuriensis</i>	45.5	33.7	0.74	B	Lo
<i>Cipangopaludina_chinensis</i>	52.6	39.3	0.75	A	Lo
<i>Cipangopaludina_japonica</i>	67.5	45.6	0.68	A	Lo
<i>Cipangopaludina_wisseli</i>	63.9	50.9	0.80	A	Le
<i>Filopaludina_bengalensis</i>	24.1	17.2	0.71	A	Lo
<i>Filopaludina_decipiens_javanica</i>	25.6	18.2	0.71	A	Le/Lo ¹
<i>Filopaludina_doliaris</i>	16.1	11.8	0.73	A	Le
<i>Filopaludina_filosa</i>	20.4	14.7	0.72	A	Lo
<i>Filopaludina_javanica</i>	24.8	18.3	0.74	A	Le/Lo ¹
<i>Filopaludina_luzonica</i>	29.1	21.6	0.74	B	Le
<i>Filopaludina_martensi</i>	33.7	28.4	0.84	B	Lo
<i>Filopaludina_polygramma</i>	20.7	15.8	0.76	A	Lo
<i>Filopaludina_sumatrensis</i>	18.6	13.9	0.75	A	Le/Lo ¹
<i>Filopaludina_tricostata</i>	27.5	20.0	0.73	B	Le
<i>Heterogen_longispira</i>	70.7	45.5	0.64	B	Le
<i>Idiopoma_sp</i>	22.0	15.9	0.72	A	Lo
<i>Larina_cf_strangei</i>	11.6	10.9	0.94	A	Lo
<i>Larina_lirata</i>	13.2	12.6	0.95	B	Lo
<i>Lioplax_cyclostomaformis</i>	19.0	12.3	0.65	A	Lo
<i>Lioplax_subcarinata</i>	19.1	12.8	0.67	A	Lo
<i>Margarya_melanioides</i>	48.0	32.0	0.67	C	Le
<i>Margarya_oxytropoides</i>	54.2	40.5	0.75	C	Le
<i>Mekongia_rattei</i>	24.0	18.0	0.75	A	Lo
<i>Mekongia_sp</i>	32.6	27.8	0.85	A	Lo
<i>Neothauma_tanganyicense</i>	26.2	22.0	0.84	A	Le
<i>Notopala_ampullaroides</i>	22.6	19.0	0.84	A	Lo
<i>Notopala_essingtonensis</i>	19.5	16.1	0.83	A	Lo
<i>Notopala_sp1</i>	20.4	16.9	0.83	A	Lo
<i>Notopala_sp2</i>	18.1	14.2	0.78	A	Lo
<i>Rivularia_auriculata</i>	24.2	16.5	0.68	A	Le
<i>Sinotaia_aeruginosa</i>	22.0	16.0	0.73	A	Le
<i>Sinotaia_quadrata</i>	29.2	18.6	0.64	A	Le/Lo ¹
<i>Sinotaia_sp</i>	25.9	16.9	0.65	B	Lo
<i>Taia_sp</i>	31.2	20.3	0.65	C	Le
<i>Tchangmargarya_yangtsunghaiensis</i>	50.2	34.6	0.69	C	Le
<i>Torotaia_cf_gilliana</i>	40.8	28.6	0.70	B	Le
<i>Torotaia_cf_lanaonis</i>	32.1	24.2	0.75	B	Le
<i>Torotaia_cf_mainitensis</i>	41.2	31.1	0.75	C	Le
<i>Torotaia_cf_mearnsi</i>	43.2	32.5	0.75	B	Le
<i>Torotaia_cf_mindanensis</i>	44.4	32.4	0.73	A	Le
<i>Trochotaia_trochoides</i>	19.9	21.7	1.09	B	Le
<i>Tulotoma_magnifica</i>	32.2	28.3	0.88	C	Lo
<i>Viviparus_ater</i>	37.0	27.6	0.75	A	Le
<i>Viviparus_cf_contectus</i>	32.0	26.0	0.81	A	Le
<i>Viviparus_georgianus</i>	22.8	20.3	0.89	A	Lo
<i>Viviparus_subpurpureus</i>	19.5	15.5	0.79	A	Lo
<i>Viviparus_viviparus</i>	36.2	26.7	0.74	A	Lo
[†] <i>Viviparus_langtonensis</i> (CP1)	–	–	–	A	Le/Lo ² Le/Lo ²
[†] <i>Campeloma_harlowtonense</i> (CP2)	–	–	–	A	Le/Lo ² Lo ²
[†] <i>Margarya_nanningensis</i> (CP3)	–	–	–	C	Le/Lo ² Le ²
<i>Bellamya cf. unicolor</i> (CP4)	–	–	–	A	Le/Lo ² Lo ²
[†] <i>Neothauma_hattinghi</i> (CP5)	–	–	–	B	Le/Lo ² Le/Lo ²

¹ species found in both habitat types, ² ambiguous; see also ‘Remarks on Habitat Types for the Five Fossils Used to Time-Calibrate the Phylogeny’ above.

TABLE S8. AIC comparison of shell-habitat estimation based on the transition models shown in Fig. S9. Model specifications: ‘HabDep’ = habitat-dependent, ‘HabInd’ = habitat-independent, ‘Hab’ = habitat transition rate, ‘Shell’ = shell transition rate, ‘ARD’ = all rates different, ‘ER’ = equal rates, and ‘SYM’ = symmetric rates.

Analysis	Model	LnL	Nparams	AIC	ΔAIC
unconstrained	HabDep-Hab_{ER}-Shell_{ARD}	-97.7	13	221.4	0.0
	HabDep-Hab _{ARD} -Shell _{ARD}	-97.1	14	222.2	0.8
	HabDep-Hab _{ER} -Shell _{SYM}	-104.9	7	223.9	2.5
	HabDep-Hab _{ARD} -Shell _{SYM}	-104.4	8	224.9	3.5
	HabInd-Hab _{ER} -Shell _{ARD}	-106.6	7	227.2	5.8
	HabInd-Hab _{ARD} -Shell _{ARD}	-105.8	8	227.5	6.1
	HabDep-Hab _{ER} -Shell _{ER}	-111.5	3	228.9	7.6
	HabDep-Hab _{ARD} -Shell _{ER}	-110.9	4	229.9	8.5
	HabInd-Hab _{ER} -Shell _{SYM}	-113.0	4	234.0	12.6
	HabInd-Hab _{ARD} -Shell _{SYM}	-112.1	5	234.3	12.9
	HabInd-Hab _{ER} -Shell _{ER}	-119.8	2	243.7	22.3
	HabInd-Hab _{ARD} -Shell _{ER}	-119.0	3	244.0	22.6
fossil-constrained	HabDep-Hab_{ER}-Shell_{ARD}	-105.3	13	236.6	0.0
	HabDep-Hab _{ARD} -Shell _{ARD}	-104.7	14	237.4	0.8
	HabInd-Hab _{ER} -Shell _{ARD}	-114.0	7	242.1	5.5
	HabInd-Hab _{ARD} -Shell _{ARD}	-113.2	8	242.5	5.9
	HabDep-Hab _{ER} -Shell _{SYM}	-114.8	7	243.6	7.0
	HabDep-Hab _{ARD} -Shell _{SYM}	-114.0	8	243.9	7.3
	HabDep-Hab _{ARD} -Shell _{ER}	-121.1	4	250.2	13.6
	HabDep-Hab _{ER} -Shell _{ER}	-122.1	3	250.2	13.6
	HabInd-Hab _{ER} -Shell _{SYM}	-123.9	4	255.8	19.2
	HabInd-Hab _{ARD} -Shell _{SYM}	-123.1	5	256.1	19.5
	HabInd-Hab _{ER} -Shell _{ER}	-131.1	2	266.3	29.7
	HabInd-Hab _{ARD} -Shell _{ER}	-130.3	3	266.6	30.0
fossil-constrained (uncertain habitats)	HabDep-Hab_{ER}-Shell_{ARD}	-106.6	13	239.2	0.0
	HabDep-Hab _{ARD} -Shell _{ARD}	-105.9	14	239.9	0.7
	HabInd-Hab _{ER} -Shell _{ARD}	-114.4	7	242.7	3.6
	HabInd-Hab _{ARD} -Shell _{ARD}	-113.5	8	243.0	3.9
	HabDep-Hab _{ER} -Shell _{SYM}	-116.0	7	246.0	6.9
	HabDep-Hab _{ARD} -Shell _{SYM}	-115.2	8	246.5	7.3
	HabDep-Hab _{ER} -Shell _{ER}	-123.3	3	252.5	13.3
	HabDep-Hab _{ARD} -Shell _{ER}	-122.4	4	252.7	13.6
	HabInd-Hab _{ER} -Shell _{SYM}	-124.2	4	256.4	17.2
	HabInd-Hab _{ARD} -Shell _{SYM}	-123.4	5	256.7	17.5
	HabInd-Hab _{ER} -Shell _{ER}	-131.4	2	266.9	27.7
	HabInd-Hab _{ARD} -Shell _{ER}	-130.6	3	267.2	28.0

TABLE S9. Transition rates obtained for the best-fit models (see Table S6 for details). Elevated transition rates are marked in grey, transition rates marked with a ‘-’ were not allowed *a priori*. Character states: ‘LeA’ = lentic with shell type A (‘absent’), ‘LeB’ = lentic with shell type B (‘fine’), ‘LeC’ = lentic with shell type C (‘coarse’), ‘LoA’ = lotic with shell type A, ‘LoB’ = lotic with shell type B, and ‘LoC’ = lotic with shell type C.

Analysis	Transition rates						
	LeA	LoA	LeB	LoB	LeC	LoC	
unconstrained HabDep-Hab_{ER}-Shell_{ARD}	LeA	-7.39	0.04	7.35	-	0	-
	LoA	0.04	-0.05	-	0.01	-	0
	LeB	8.78	-	-8.88	0.04	0.06	-
	LoB	-	0.13	0.04	-0.17	-	0
	LeC	0	-	0.02	-	-0.06	0.04
	LoC	-	0	-	0.28	0.04	-0.32
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	fossil-constrained HabDep-Hab_{ER}-Shell_{ARD}	LeA	-4.05	0.03	4.02	-	0
LoA		0.03	-0.06	-	0.03	-	0
LeB		4.64	-	-4.75	0.03	0.08	-
LoB		-	0.26	0.03	-0.29	-	0
LeC		0	-	0.03	-	-0.07	0.03
LoC		-	0	-	0.27	0.03	-0.31
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fossil-constrained (uncertain habitat) HabDep-Hab_{ER}-Shell_{ARD}		LeA	-8.36	0.04	8.32	-	0
	LoA	0.04	-0.06	-	0.02	-	0
	LeB	10.00*	-	-10.11	0.04	0.07	-
	LoB	-	0.23	0.04	-0.27	-	0
	LeC	0	-	0.03	-	-0.07	0.04
	LoC	-	0	-	0.25	0.04	-0.29

* Note that the maximum value for rates was fixed to 10 to fit the models in the fossil-constrained analyses.

REFERENCES

- Doello-Jurado M. 1922. Una especie de “Viviparus” del cretáceo superior de Rio Negro. *Physis Buenos Aires* 5:328–330.
- Etheridge R. 1902. A monograph of the Cretaceous invertebrate fauna of New South Wales. *Mem. Geol. Surv. New South Wales* 11:1–98.
- Gerstfeldt G. 1859. Über Land- und Süßwasser-Mollusken Sibiriens und des Amur-Gebietes. *Mémoires présentés à l’Académie Impériale des Sci. St.-petersbg. par Divers savants lus dans ses Assem.* 9:505–548.
- Gill T. 1863. Systematic arrangement of the mollusks of the family Viviparidae, and others, inhabiting the United States. *Proc. Acad. Nat. Sci. Philadelphia* 15:33–40.
- Hartman J.H., Erickson D.N., Bakken A. 2008. Stephen Hislop and his 1860 Cretaceous continental molluscan new species descriptions in Latin from the Deccan Plateau, India. *Palaontology* 51:1225–1252.
- Hirano T., Saito T., Chiba S. 2015. Phylogeny of freshwater viviparid snails in Japan. *J. Molluscan Stud.* 81:435–441.
- Hislop S. 1860. On the Tertiary deposits, associated with trap-rock, in the East Indies – with descriptions of the fossil shells. *Q. J. Geol. Soc. London* 16:154–182, 188, 189.
- Hudleston W.H. 1887-1896. A monograph of the inferior Oolite Gasteropoda. Being part I of the British Jurassic Gasteropoda. London: Palaeontological Society London.
- Kear B.P., Hamilton-Bruce R.J., Smith B.J., Gowlett-Holmes K.L. 2003. Reassessment of Australia’s oldest freshwater snail, *Viviparus* (?) *albascopularis* Etheridge, 1902 (Mollusca: Gastropoda: Viviparidae), from the Lower Cretaceous (Aptian, Wallumbilla Formation) of White Cliffs, New South Wales. *Molluscan Res.* 23:149–158.
- Nevill G. 1877. List of the mollusca brought back to Mr. Anderson from Yunnan and upper Burma with descriptions of new species. *J. Asiat. Soc. Bengal* 46:14–41.
- Philippi R.A. 1887. Die Tertiären und Quartären Versteinerungen Chiles. Leipzig: F.A. Brockhaus.
- Pickford M. 2004. Palaeoenvironments of Early Miocene hominoid-bearing deposits at Napak, Uganda, based on terrestrial molluscs. *Ann. Paléontologie* 90:1–12.
- Prashad B. 1928. Recent and fossil Viviparidae. A study in distribution, evolution and palaeogeography. *Mem. Indian Museum* 8:153–251.
- Quan C., Fu Q., Shi G., Liu Y., Li L., Liu X., Jin J. 2016. First Oligocene mummified plant Lagerstätte at the low latitudes of East Asia. *Sci. China Earth Sci.* 59:445–448.
- Schultheiß R., Van Bocxlaer B., Riedel F., von Rintelen T., Albrecht C. 2014. Disjunct distributions of freshwater snails testify to a central role of the Congo system in shaping biogeographical patterns in Africa. *BMC Evol. Biol.* 14:42.
- Scotese C.R. 2014a. Atlas of Neogene Paleogeographic Maps (Mollweide Projection), Maps 1-7, Volume 1, The Cenozoic, PALEOMAP Atlas for ArcGIS, PALEOMAP Project, Evanston, IL.
- Scotese C.R. 2014b. Atlas of Paleogene Paleogeographic Maps (Mollweide Projection), Maps 8-15, Volume 1, The Cenozoic, PALEOMAP Atlas for ArcGIS, PALEOMAP Project, Evanston, IL.
- Scotese C.R. 2014c. Atlas of Early Cretaceous Maps (Mollweide Projection), Maps 23-31, Volume 2, The Cretaceous, PALEOMAP Atlas for ArcGIS, PALEOMAP Project, Evanston, IL.
- Scotese C.R. 2014d. Atlas of Jurassic Paleogeographic Maps (Mollweide Projection), Maps 32-42, Volume 4, The Jurassic and Triassic, PALEOMAP Atlas for ArcGIS, PALEOMAP Project, Evanston, IL.
- Sengupta M.E., Kristensen T.K., Madsen H., Jorgensen A. 2009. Molecular phylogenetic investigations of the Viviparidae (Gastropoda: Caenogastropoda) in the lakes of the Rift Valley area of Africa. *Mol. Phylogenet. Evol.* 52:797–805.
- Simone L.R.L. 2004. Comparative morphology and phylogeny of representatives of the superfamilies of architaenioglossans and the Annulariidae (Mollusca, Caenogastropoda). *Arq. do Mus. Nac. Rio Janeiro* 62:387–504.
- Stanton T.W. 1903. A new fresh-water molluscan faunule from the cretaceous of Montana. *Proc. Am. Philos. Soc.* 42:188–199.
- Thompson F.G. 1999. An identification manual for the freshwater snails of Florida. *Walkerana*. 10:1–96.
- Tian Y., Shaw D., Schneider S. 2018. Oligocene fossil assemblages from Lake Nanning (Yongning Formation; Nanning Basin, Guangxi Province, SE China): biodiversity and evolutionary implications. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 505:100–119.
- Tracey S., Todd J.A., Erwin D.H. 1993. Mollusca: Gastropoda. In: Benton M.J., editor. *The fossil record 2*. London: Chapman and Hall. p. 131–167.
- Van Bocxlaer B., Strong E.E. 2016. Anatomy, functional morphology, evolutionary ecology and systematics of the invasive gastropod *Cipangopaludina japonica* (Viviparidae: Bellamyinae). *Contrib. to Zool.* 85:235–263.
- Van Damme D., Pickford M. 1999. The late Cenozoic Viviparidae (Mollusca, Gastropoda) of the Albertine Rift Valley (Uganda-Congo). *Hydrobiologia* 390:171–217.
- Yen T.-C. 1950. Fresh-water mollusks of Cretaceous age from Montana and Wyoming. *Geol. Surv. Prof. Pap.* 233-A:1–20.
- Zhang L.-J., Chen S.-C., Yang L.-T., Jin L., Köhler F. 2015. Systematic revision of the freshwater snail *Margarya* Nevill, 1877 (Mollusca: Viviparidae) endemic to the ancient lakes of Yunnan, China, with description of new taxa. *Zool. J. Linn. Soc.* 174:760–800.